AN EXPOSITION ON SPECTROPHOTOMETRIC DETERMINATION OF PLATINUM GROUP METALS

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ABSTRACT

In the realm of analytical chemistry, transition metal complexes especially the platinum group metals (PGMs) are often used compounds. Strong chromophores, inflexible luminous structures, biological potential and electrochemical activity are characteristics of the platinum metal complexes. These features have prompted the creation and study of analytical techniques based on these metal complexes for the identification of different analytes in a variety of analytical applications. Even if there are a few broad recommendations accessible to forecast a reagent's potentialities for the given goal, these studies show that sensitivity and selectivity imparted by the reagent to the metal in respective complex must be established. The quest for novel reagents is an ongoing effort to the wide and varied need for new ways to identify and analyze the metal ions under the peculiar conditions. In light of the frightening and complicated issue of environmental contamination, the endeavor of developing new and innovative reagents as well as techniques for inorganic analysis of PGMs have particular relevance. Of the numerous methodologies, the UV/VIS spectrophotometric determination technique of the PGM complexes is of interest being reliable, easy to handle, quick, cost effective, selective and sensitive. The technique relies on the process of interaction between an analyte and the metal, which are then determined analytically. In light of the enrichment of platinum group metals, the purpose of this article is to outline current research in the methods of spectrophotometric determination of PGMs. The review will assist researchers in formulating and refining PGMs as workable candidates for applications in medicine, pharmacology, analysis and catalysis.

Keywords: Platinum Group Metals; Platinum metals; PGMs; Extraction; Spectrophotometric determination.

1. INTRODUCTION

The PGMs in form of their compounds/complexes are significant in several domains due to their distinct coordination chemistry. The Platinum Group Metals, also called platinoids, platidises, platinides, platinum group, platinum metals or platinum family, are a family of six elements *viz*. Ruthenium, Rhodium, Palladium, Osmium, Iridium and Platinum belonging to groups 8, 9 and 10 in the periodic table of elements that are structurally and chemically similar and known for their array of applications including industrial, medicinal and electronic. These are imperative noble metals known for their distinct chemical and physical properties [1-3]. Depending on their atomic numbers (or weights), the PGMs metals are sometimes divided into light platinum group metals (Ru, Rh, Pd) and heavy platinum group metals (Os, Ir, Pt) [4]. These can also be sub-divided as iridium-group platinum group elements (osmium, iridium and ruthenium) and the palladium-group platinum group elements (rhodium, platinum, palladium) based upon their behaviour in geological systems.

1.1 Discovery of PGMs

The English Scientist William Hyde Wollaston was the first to isolate pure platinum, palladium and rhodium in 1803-04; Smithson Tennant found pure iridium in 1803, as did N.Z. Vauquelin and A.F. de Fourcroy in France. In 1803 or 1804, Tennant also discovered osmium. The Russian chemist K.K. Klaus made the latest PGM, ruthenium, to be isolated in 1844. Palladium, rhodium and iridium are all named after their respective Greek origins. Palladium was given its name in honor of the asteroid, Pallas, since both objects were discovered about the same time. Rhodium receives its name from the hue of aqueous, acidic solutions of metal salts, which is taken from the word, rhodon, meaning rose. Osmium is derived from osme, which means scent, due to the unique aroma of its volatile tetraoxide and iridium comes from the term iris, rainbow like, due to the vast spectrum of colors that iridium salt solutions show. Ruthenium is named after Ruthenia, the Latin name for Russia and the last PGM, the Platinum which was first found in the sixteenth century in Columbia's Choco district, found its name originated from the word, platina in Spanish, meaning little silver [5-7].

1.2 The Principal PGM Sources

PGMs, low-prevalence elements, are primarily supplied by South Africa, with reserves estimated at 63,000 tons in 2020, followed by Russia, Zimbabwe, the US and Canada. Platiniridium is the naturally occurring platinum-iridium alloy; however, osmiridium is the known natural alloy of iridium and osmium. The

platinum group metals can be either sulphur-loving or iron-loving and can also be pronounced as chalcophiles and siderophores, respectively. The common minerals of PGMs are-Tellurides, selenides, arsenides and antimonides. There are several significant sulphide minerals of Ru, Pd and Pt. The PGM deposits are indicated in Table 1 [8], where the deposits in Merenskey Reef are seen associated with iron, nickel and copper ores. The major source of the metals is Sudbury area in Canada where the metals are found as by-product of nickel and copper production. However, in general, the platinum group metals are known to exist together with their ratio variable with the ore location [9, 10].

1.3 Advantages of PGMs

Six noble metals with a white-silver hue make up the platinum group: ruthenium, iridium, osmium, palladium, platinum and rhodium. PGMs are grouped collectively in the periodic table of elements based on similar mineralization and characteristics. Without a question, the most well-known PGM is platinum due to its thermal stability as well as melting point [11]. The metal is compatible with biological systems and acts as an effective oxidation catalyst. Palladium is less common than platinum, yet it is nonetheless useful for a variety of chemical processes because of its special capacity to absorb hydrogen. With a high melting point and exceptional thermal stability, rhodium is also known to possess good catalytic properties . Iridium is an extremely rare metal that is resistant to corrosion. It is also highly stable both chemically and thermally, dense and compatible with life. Ruthenium possesses strong melting point and is an excellent alloying agent exhibiting good conductive and catalytic qualities. The densest and hardest metal, osmium is frequently used in combination with other PGMs and is another efficient catalyst and superb electrical conductor [1, 12, 13].

Table 1: Platinum Group Metal Content (grams tonne⁻¹) in Ore Bodies

Location of PGMs Deposits	Ruthenium g/t	Rhodium g/t	Palladium g/t	Osmium g/t	Iridium g/t	Platinum g/t	Total PGM g/t			
Canada										
Sudbury Complex	0.04	0.03	0.4	0.01	0.01	0.3	0.79			
Lac-des lles	-	-	2.3	-	-	0.2	2.5			
	'	U	SA	•	•	1				
Stillwater Complex	0.36	0.6	11	-	0.21	3.3	15.47			
		Zimb	oabwe			ı				
Hartely Complex	-	0.21	1.8	-	-	2.6	4.61			
	Russia									
Noril'sk	-	0.24	-	-	-	2.5	9.74			
		South	Africa							
Merensky Reef	0.33	0.16	1.4	0.04	0.05	2.7	4.68			
-East Bushveld	0.23	0.13	1.22	-	0.04	2.32				
-West Bushveld	0.63	0.28	1.6	-	0.14	3.6				
Platreef	0.14	0.12	1.9	0.03	0.04	1.9	4.13			
-North Bushveld	-	0.08	1.1	-	-	0.82	2.0			
UG2	0.45	0.34	1.3	0.05	0.13	2.0	4.27			
-East Bushveld	0.76	0.45	2.06	-	0.18	2.42				
-West Bushveld	0.93	0.54	1.48	-	0.22	2.89				

The Platinum group elements show a diverse array of applications in the following industries: electronics, gas remediation, automotive industry, petroleum refining, fine chemical and ammonia manufacturing and the environment [8, 14]. However, the principal application of platinum group metals is in alloys and as catalysts in numerous industrial manufacturing. The PGMs being a part of luxurious lifestyle are used in jewellery making because of their property of thermal stability and corrosion-resistance [15]. The medical field is highly enlightened with the application of PGMs [16-19]. The metal complexes have received a lot of interest recently from the medical world and from therapeutics treating terrible diseases like cancer due to the metals' propensity for organic substrates, their activity in redox reactions and their ability to exist in a variety of oxidation states. Because of the mentioned characteristics of metal complexes, platinum group based complexes have occasionally been used in the remedial field, giving them notable qualities in the subsequent field [20-22].

The wide range of applications for platinum group metals based complexes, including industrial, chemical, catalytic and electronic, as well as the potential to treat deadly diseases like cancer, make them highly valuable resources that require careful consideration and diligence to advance both the prosperity of these fields and the quality of healthcare in general [8]. In this context, attempts have occasionally been made to develop methods and procedures pertinent to the creation of novel compounds of the metals and also the protocols for figuring out their trace concentrations and hence the micro scale determination.

2. Determination of Platinum Group Metals

Trace analysis of metals has evolved significantly since World War II, driven by the need for high purity materials. It is a valuable tool in various industries, including food, environmental pollution and archeological research [23]. Trace amounts of metals and nonmetals are essential for biological processes as well, but toxic concentrations can pose a threat. In light of the variety and unique

complexity in the structures of PGM compounds as well as the chemical changes they undergo in both natural and industrial systems, the coordination and analytical chemistry of the platinum metals is a distinct subject in chemistry. Trace element determination can be done using a variety of instrumental techniques. Among the most suitable analytical techniques that have been suggested thus far are gravimetric [24, 25], titrimetric [26], AAS [27], X-ray analysis [28, 29], nuclear [30, 31], potentiometry and voltammetry [32], inductively coupled plasma atomic emission spectrometry (ICP-AES) [33], inductively coupled plasma mass spectrometry (ICP-MS) [34], electrothermal atomic absorption spectrometry (ET-AAS) [35] and UV-VIS spectrophotometry [36]. Nonetheless, the expediency of the aforementioned methods for the direct determination of platinum metals is compromised by a number of shortcomings and deficiencies, such as cost-ineffectiveness, safety concerns and maintenance challenges. Therefore, it becomes necessary to create and discover a methodology that can withstand and overcome all of the shortcomings of the aforementioned approaches [37, 38]. Spectrophotometry is the most commonly used technique for trace metal determination since the very beginning, as it is simple and inexpensive method. Using appropriate purity reagents is crucial for sensitive spectrophotometric techniques. UV/VIS spectrophotometry, hence, has developed into a powerful analytical tool to address the demerits of the current methodologies.

Spectrophotometric Methods of Determination of PGMs

When compared to other applied analytical techniques, various organic reagents/ligands have been noted to be sensitive, cost-effective, precise and accurate when utilized for the trace level determination of the platinum metals employing a UV/VIS spectrophotometric methodology. As a result, a thorough and concise analysis of the existing spectrophotometric techniques is carried out on PGMs and provided below to determine the extent to which these methods satisfy the mentioned criteria.

2.1 Thiourea derivatives

Thiourea (Figure 1) and its derivatives have unique structural and electrical properties, making them useful in biological and non-biological sectors. These form hydrogen bonds and offer coordination sites as ligands, making them useful as chemosensors for identifying metal cations in biological, agricultural and environmental samples. Owing to its great sensitivity, selectivity and cheap cost of synthesis, thiourea and its derivatives are used as organic chemosensors, producing stable complexes with numerous cations including those with PGMs that are vital to different fields [39]. The spectrophotometric aspects of the different derivatives (Figure 2) have been collected together in the Tables 2a to 2d as below:

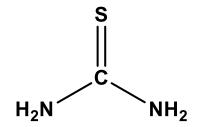


Figure 1: Thiourea

Table 2a: Spectrophotometric Determination of PGMs using Thiourea Derivatives as Ligands

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
Ethylene thiourea (ETU)	Os (VIII)	1M H ₂ SO ₄ , pH 1.0; 1:3	Water; 490	16.87 x 10 ⁴	Interferences of few ions masked using EDTA and fluoride	40
Ethyl thiourea	Os (III)	HCl; 2%NH4[Cr(CNS)4(aniline)], acetone; 1:3	Acetone; 535-540	793.067	Less sensitive	41
N,N'-Diphenylbenzimidoyl Thiourea (DPB1TU)	Pd (II)	0.5M HCl; equilibration time 1 min.; 1:2	CHCl ₃ ; 365	2.50 x 10 ⁴	1 min. extraction time	42
N-octyl-N'-(sodium p- amminobenzenesulfonate)- thiourea (OPT)	Pd (II)	Buffer, pH 5.2, Heating at 50°C for 6 min., CTMAB; 1:4	Water; 299	1.38 x 10 ⁵	Heating at 50°C for 6 min	43
N-(3,5-Dimethylphenyl)-N'-(4- Aminobenzene sulfonate) thiourea ((DMMPT)	Pt (IV)	Buffer, pH 3.8, colour development time 10 min., CTMB; 1:3	CH ₂ Cl ₂ ; 755	9.51 x 10 ⁴	Colour development time 10 min.	44
N-(3,5-Dihydroxylphenyl)-N'- (4-Aminobenzene sulfonate) thiourea (DHPABT)	Pt (IV)	5M HCl, Colour development time 10 min.; 1:2	Water; 760	1.01 x 10 ⁵	Colour development time 10 min.	45
N-dodecyl-N'-(sodium-p- aminobenzenesulfonate) thiourea (DOPT)	Pd (II)	Buffer, pH 5.4, Heating at 100°C, CTMB; 1:4	Water; 296.4	7.41 x 10 ⁴	Heating at 100°C required; Cu ²⁺ interferes	46
N-(m-Methylphenyl)-N'- (sodium p- aminobenzenesulfonate)- thiourea (MMPT)	Pt (IV)	Buffer, pH 4.0, Heating at 90° C for 3min.; 1:3	Water; 754.4	8.58 x 10 ⁴	Heating at 90° C for 3 min.	47

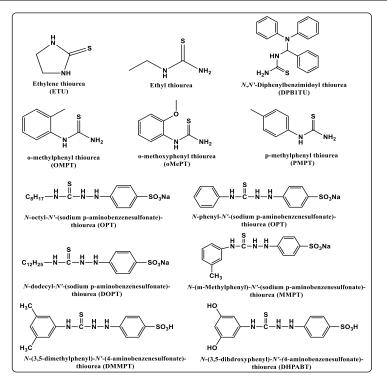


Figure 2: Thiourea derivatives as chromogenic ligands

Table 2b: Spectrophotometric Determination of PGMs using Thiourea Derivatives as Ligands

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
	Pd (II)	0.8M HCl, 5 sec. equilibration time; 1:1	CHCl ₃ ; 340	2.85 x 10 ³	Ag (I) interferes	48
	Pt (IV)	0.1M KI; 1:1:2	CHCl ₃ ; 362	1.25 x 10 ⁴	-	49
o-methylphenylthiourea (OMPT)	Ru (III)	3M HCl; Heating for 5 minutes; 1:2	CHCl₃; 590	2.34 x 10 ³	Heating on boiling water bath for 5 minutes	50
	Os (IV)	0.8M HCl; 1:1	CHCl ₃ ; 514	1.86 x 10 ³	Pd (II), Ir (III) interfere	51
	Rh (III)	Buffer, pH 5.4, Heating for 4 minutes; 1:2	CHCl₃; 320	9.76 x 10 ³	Heating in boiling water bath for 4 minutes	52
	Ir (III)	0.6M HCl; 1:1	CHCl ₃ ; 516	0.663 x 10 ³	Pd (II), Os (VIII) interfere	53

Table 2c: Spectrophotometric Determination of PGMs using Thiourea Derivatives as Ligands

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (L mol ⁻¹ cm ⁻¹)	Comments	Reference
o-methoxyphenyl thiourea (OMePT)	Pd (II)	1M HCl; 1:1	CHCl ₃ ; 325	3.38 x 10 ³	Ag (I) interferes	54
	Os (IV)	0.8M HCl; 1:1	CHCl ₃ ; 518	2.12×10³	Pd (II), As (III), Ir (III) interferes	55
	Ru (III)	3.4M HCl,Heating for 5 minutes; 1:2	CHCl ₃ ; 640	2.34 ×10 ³	Heating in boiling water bath for 5 minutes; Ir (III) interferes	55
N-phenyl-N'-(sodium p- aminobenzenesulfonate)-thiourea (PPT)	Pt (IV)	Buffer, pH 4.0, CTMAB,5 min. heating at 80°C; 1:3	Water; 778.4	2.95 x 10 ⁴	5 min. heating at 80°C in water bath; Cu (II) interferes	56
	Pd (II)	Buffer,pH 5.5, CTMAB,5 min. heating at 80°C; 1:4	Water; 306.1	7.98 x 10 ⁴	5 min. heating at 80°C in water bath; interference was observed by Cu²+	56

Table 2d: Spectrophotometric Determination of PGMs using Thiourea Derivatives as Ligands

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
	Ru (III)	0.2M HBr, heating for 5min. at 70° C; 1:1	CHCl ₃ ; 600	2.31 x 10 ³	5min. heating in boiling water bath at 70° C	57
	Ir (III)	0.1M HClO ₄ ; 1:1	CHCl ₃ ; 482	0.909 x 10 ³	Pd(II) requires prior extraction	58
p-methylphenyl thiourea	Os (VIII)	0.45M HClO ₄ ; 1:1	CHCl ₃ ; 512	6.83 x 10 ³	5 sec equilibration time	59
(PMPT)	Pt (IV)	0.03M KIO ₃ ; 1:1	CHCl ₃ ; 506	6.78 x 10 ³	Os (VIII) and Ir (III) requires prior extraction	60
	Pd (II)	0.05M HBr,10 sec. equilibration time; 1:1	CHCl ₃ ; 300	8.486 x 10 ³	10 sec equilibration time	61

2.2 Thiosemicarbazones derivatives

Thiosemicarbazones (Figure 3) are employed as a chelating ligand in the production of metal complexes because of their adaptable donor sets, including nitrogen and sulphur [62]. These can coordinate in a variety of ways to produce coordination compounds with different metal ions. It is a condensed mixture of ketone or aldehyde and thiosemicarbazone [63]. Because of their bonding mechanisms and the presence of amide, imine and thione groups, which make them potential polydentate ligands, thio and phenyl thiosemicarbazones have drawn increasing interest in recent years [64]. Behaving as anti-retroviral, anti-microbial, anti-tubercular, anti-HIV, anti-fungal, anti-trypanosomal, anti-amoebic, anti-malarial, anti-viral, anti-tumor, anti-bacterial and cytotoxic substances, are a few of the biological characteristics of thiosemicarbazones. Due to their particular and selective coordination capabilities, thiosemicarbazones themselves have the potential to be used as metal ion for sensing and scavenging of metals. Their metal complexes have also been identified to have potential use

in medical imaging [65]. The till date investigated PGM complexes using thiosemicarbazones as ligands (Figure 4) have been put under tables 3a-3d.

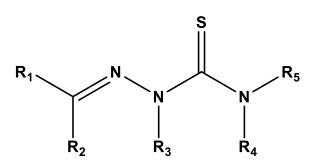


Figure 3: Thiosemicarbazones

Table 3a: Spectrophotometric Determination of PGMs using Thiosemicarbazone derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
p-[N,N-bis(2-chloroethyl) amino]Benzaldehyde thiosemicarbazone (CEABT)	Pd (II)	0.2M HCl, CEABT; 1:2	Water; 395	4.05 x 10 ⁴	Cu(II) and Pt(VI) interfere	66
4-[<i>N</i> , <i>N</i> -(diethyl)amino] benzaldehyde thiosemicarbazone (DEABT)	Pt (IV)	Buffer, pH 3.0; 1:2	Water; 405	1.755 x 10 ⁴	Cu(II) and Pd(II) interfere	67
Phthalaldehydic acid thiosemicarbazone (PAATSC)	Pd (II)	2M Buffer , pH 4.0, 1min. equilibration time; 1:10	C ₆ H ₁₂ O; 355	5.1 x 10 ⁴	1min. equilibration time	68
Benzildithiosemicarbazone (BDTSC)	Pd (II)	Buffer, pH 2.5, 30 sec. equilibration time 1:1	(CH ₃) ₂ CHCH ₂ CCH; 395	3.018 x 10 ⁴	Ag(I) requires prior extraction	69
2,6-Diacetylpyridine bis-4-phenyl-3-thiosemicarbazone (2, 6-DAPBPTSC)	Pd (II)	Buffer, pH 4.0, 1min. equilibration time; 1:1	CsH12O; 410	1.156 x 10 ⁴	1min. equilibration time	70
Isonitroso p-nitroaceto phenone thiosemicarbazone (HINATS)	Pd (II)	1M CH ₃ COOH, 1min. equilibration time; 1:1	CHCl ₃ ; 410	910	1min. equilibration time, Less sensitive	71
Pyridoxalthiosemicarbazone (PTSC)	Pd (II)	Buffer , pH 2.0; 1:2	Water; 420	1.63 x 10 ⁴	-	72
N-Ethyl-3-carbazole carbaxaldehyde thiosemicarbazone (ECCT)	Pd (II)	Buffer, pH 4.0, 3 min. equilibration time; 1:1	C ₄ H ₉ OH; 410	1.647 x 10 ⁴	3 min. equilibration time	73
3,4-dihydroxy-5-methoxy Benzaldehyde thiosemicarbazone (DHMBTSC)	Ru (III)	Buffer , pH 2.0, Triton X-100; 1:1	Water; 425	2.73 x 10 ⁴	-	74

Table 3b: Spectrophotometric Determination of PGMs using Thiosemicarbazone derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
Phenanthraquinone monothiosemicarbazone	Ru (III)	Buffer, pH 5.2-6.7, Heating for 2 hours; 1:2	Water; 660	9843.81	Heating in water bath for 2 hours, Cu(II) and Rh(III) interfere	75
	Rh (III)	0.2M C ₈ H ₅ KO ₄ , Heating for 90 min.; 1:2	Water; 510	7900	90 minutes heating on water bath is required, Ru(III), Pd(II) and Cu(II) interfere	76
(PTS)	Os (VIII)	CH ₃ OH, pH 6.8-8.5; 1:3	Water; 530	15280	Ni(II), Cu(II), Co (II), Ru (III), Rh (III) and Ir(III) interfere	77

Table 3c: Spectrophotometric Determination of PGMs using Thiosemicarbazone derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; $\lambda_{max}(nm)$	Molar Absorptivity (L mol ⁻¹ cm ⁻¹)	Comments	Reference
	Ru (III)	Buffer, pH 4.6, Digested for 25min., Equilibration time 1 min.; 1:3	CH ₃ (CH ₂) ₄ OH; 505	9855.95	Digested for 25min. in boiling water bath; Equilibration time 1 min.	78
	Pd (II)	2M HCl, Equilibration time 1 min.; 1:2	CH ₃ (CH ₂) ₄ OH; 520	7450	Equilibration time 1 min.	79
2-(5- Bromo-2- oxoindolin-3-ylidene)	Os (IV)	Buffer , pH 4.5, Digested for 30 min.; 1:2	CH ₃ (CH ₂) ₄ OH; 565	10000	Digested for 30 min. in boiling water bath	80
hydrazine carbothioamide (HBITSC)	Rh (III)	Buffer , pH 5.5, Digested for 30 min., Equilibration time 1 min.; 1:3	CH ₃ (CH ₂) ₄ OH; 495	3345	Digested for 30 min. in boiling water bath; Equilibration time 1 min.	81
	Pt (IV)	Buffer, pH 4.6, Digested for 30 min., Equilibration time 1 min.; 1:2	CH ₃ (CH ₂) ₄ OH; 505	8452.53	Digested for 30 min. in boiling water bath	82
	Ir (III)	Buffer, pH 6.5, Digested for 30 min.; Equilibration time 1 min.; 1:3	CH ₃ (CH ₂) ₄ OH; 500	1153.35	Digested for 30 min. in boiling water bath; Equilibration time 1 min.	83

Table 3d: Spectro	photometric Det	termination of	f PGMs using	Thiosemicarbazone	derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
2-hydroxy-3-methoxy Benzaldehyde thiosemicarbazone (HMBATSC)	Ru (III)	Buffer, pH 2.5; 1:1	Water; 390	0.964 x 10 ⁴	-	84
	Pd (II)	Buffer, pH 3.0; 2:3	Water; 380	2.198 x 10 ⁴	-	85
Piperonal thiosemicarbazone (PATS)	Pd (II)	0.5M HCl; 1:2	Water; 363	3.80 x 10 ⁴	Cu(II),Hg(II) and Pt(II) interfere	86
	Pt (IV)	0.2M H ₂ SO ₄ ; equilibration time 10 min.; 1:2	Water; 360	3.24 x 10 ⁴	Cu(II), Hg(II),Au(III)and Pd(II) interfere	87
Pyridoxal-4- phenyl-3- thiosemicarbazone (PDPTSC)	Pd (II)	Buffer, pH 3.0, equilibration time 1 min.; 1:1	C ₆ H ₆ ; 460	2.20 x 10 ⁴	Equilibration time 1 min.	88
	Pt (IV)	Buffer , pH 3.5	CHCl ₃ ; 460	3.561 x 10 ³ - 7.641 x 10 ³	Less sensitive method	89

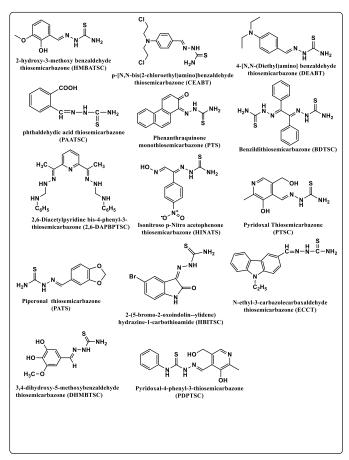


Figure 4: Thiosemicarbazone derivatives as chromogenic reagents.

2.3 Hydrazones derivatives

In inorganic chemistry, hydrazones play a critical role in building complexes with transition metal ions. These compounds include a conjugated C-N bond with a lone pair of electrons on the functional atom [90]. While the nitrogen atoms in hydrazones (Figure 5) are nucleophilic, the carbon atom behaves both electrophilic and nucleophilic. Hydrazones have special chemical and physical characteristics along with a C-N connection [91]. They can cure illnesses including TB, leprosy and mental problems while acting as models for biological species. Hydrazones serve as the versatile chemical agents, including nematocides, herbicides, insecticides, rodenticides and plant growth regulators [92]. Since hydrazones may readily form stable complexes with the majority of transition metal ions, these are useful in inorganic chemistry. Hydrazone complexes have garnered more attention with increasing growth of bioinorganic chemistry, partly because several of these complexes have been identified as potential models for biologically significant species [93]. Hydrazones have been

the subject of increasing interest in research due to their anticancer, antimicrobial and antitubercular properties [94, 95]. Analytically, the hydrazone derivatives (Figure 6) help in determination of PGMs using numerous spectrophotometric procedures, briefed as under (Tables 4a -4b).

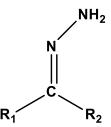


Figure 5: Hydrazones

Table 4a: Spectrophotometric Determination of PGMs using Hydrazone derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (L mol ⁻¹ cm ⁻¹)	Comments	Reference
2,4-dihydroxyl acetophenone-p- hydroxylbenzoylhydrazone (RPPHBH)	Pd (II)	Buffer, pH 5.5; 1:1	Water; 375	0.85 x 10 ⁴	-	96
Diacetyl Monoxime Isonicotinoylhydrazone (DMIH)	Ru (II)	Buffer, pH 4.5; 1:1	Water; 346	1.4 x 10 ⁴	-	97
3,5-Dimethoxy-4-hydroxy	Ru (II)	Buffer, pH 4.5; 1:1	Water; 390	1.7 x 10 ⁴	-	98
benzaldehydeisonicotinoylhydrazone (DMHBIH)	Pd (II)	Buffer, pH 5.5, Triton X-100; 1:1	Water; 382	2.2 x 10 ⁴	Cu(II), Cr(VI), Au(III) and Ru(III) seriously interfere	99
2-Aminoacetophenone isonicotinoylhydrazone (2-AAINH)	Pd (II)	Buffer, pH 4.0; 1:2	C ₃ H ₇ NO; 500	3.0 x 10 ⁴	-	100
2,4-Dimethoxy benzaldehyde isonicotinoyl hydrazone (DMBIH)	Os (VIII)	Buffer, pH 5.0, Triton X-100; 1:2	Water; 393	1.48 x 10 ⁴	-	101
5-Bromo salicylaldehyde Isonocotinoylhydrazone (5-BrSAINH)	Pd (II)	DMF, pH 2.5-3.0; 1:1	Water; 445	1.3 x 10 ⁴	Fe (III) and Hg(II) interfere	102
2-Hydroxy-5-methylaceto Phenoneisonicotinoylhydrazone (HMAINH)	Pd (II)	0.01M H ₂ SO ₄ 60-90 sec. equilibration time; 1:1	CHCl ₃ ; 385	0.5320 x 10 ⁴	Shaking time of 60s to 90s required for complete extraction	103
3,4-dihydroxybenzalde- hydeisonicotinoyl Hydrazine (3,4-DHBINH)	Pd (II)	Buffer, pH 3.0; 1:1	Water; 380	0.53 x10 ⁴	-	104
Cinnamaldehydeisonicotinoylhydrazone	Ru (III)	Buffer, pH 3.0; 1:1	Water; 402	4.56 x 10 ⁵	-	105
(CINH)	Pd (II)	Buffer, pH 3.0-6.0; 1:1	Water; 412	1.03 x 10 ⁴	Mo(VI), Cu(II), Th (IV) and V(V) interfere	106
3,5-Dimethoxy-4-hydroxy benzaldehydeisonicotinoylhydrazone (DMHBIH)	Pd (II)	Buffer, pH 5.5, Triton X-100; 1:1	Water; 382	2.44 x 10 ⁴	Cu(II), Cr(VI), Au (III) and Ru (III) seriously interfere	107
	Ru(III)	Buffer, pH 4.25, Triton X-100; 1:1	Water; 390	1.7 x 10 ⁴	V(V), Cr(VI), As (III) and Sb (III) interfere	108
4-Hydroxy-3,5-dimethoxy benzaldehyde-4- hydroxyl	Pd (II)	Buffer, pH 3.0, Triton X-100; 1:1	Water; 373	7.5 x 10 ⁴	Cu(II), Fe(III) and Mo (VI) interfere	109
benzoylhydrazone (HDMBHBH)	Ru (VI)	5% CTAB, pH 5.5; 1:1	Water; 420	3.00 x 10 ⁴	-	110
Cinnamaldehyde-4-hydroxybenzoyl hydrazine (CMHBH)	Pd (II)	Buffer (pH 4.0); 1:1	CH ₂ Cl ₂ ; 375	6.0 x 10 ⁴	Cu(II), Fe(III), Mo (VI) interfere	111
2-Hydroxy-1-naphthaldehyde-p- hydroxybenzoic hydrazine (HNHBH)	Pd (II)	Buffer, pH 4.0, Triton X-100; 1:1	Water; 430	2.48 x 10 ⁴	-	112
	Pd (II)	1M CH ₂ COOH, 2M Buffer, Heating for 4-5 min. 1:2	C ₆ H ₅ CH ₃ ; 470	1.3305 x 10 ⁴	Heating on boiling water bath for 4-5 min.	113
Isonitroso-p-methyl acetophenone phenyl hydrazone (HIMAPH)	Rh (III)	2M Buffer, pH 5.0, Heating for 50-60min.; 1:3	C ₆ H ₅ CH ₃ ; 465	2631.58	Heating on water bath 50-60 min.	114
	Ru (III)	2M Buffer, pH 4.5-5.5, Heating for 30-35min; 1:3	C ₆ H ₅ CH ₃ ; 460	3535.35	Heating for 30-35min., Less sensitive	115
	Os (VIII)	0.8M HCl, 5 min. equilibration time; 1:2	CHCl ₃ ; 440	8.94 x 10 ³	Equilibration time 5min.	116
2-nitrobenzaldehyde thiocarbohydrazone	Ru (III)	0.7M Aq.CH ₃ COOH, 5 min. heating; 1:3	C ₂ H ₂ Cl ₂ ; 445	1.41 x 10 ⁴	5 min. heating on water bath	117
(2-NBATCH)	Pt (IV)	1,4-dioxane, Heating for 5min., Equilibration time of 5 min.; 1:2	CHCl ₃ ; 440	1.03 x 10 ⁴	5 min. heating in boiling water bath; Equilibration time of 5 min.	118
5-chloro-2-hydroxythio benzhydrazide	Os (VIII)	HCl, Heating for 25 min., Waiting time 30min.; 1:2	CHCl ₃ ; 510	1.056 x 10 ⁴	Heating on boiling water bath for 25min.; Waiting time 30min.	119
(5-Cl-2-OHTBH)	Ru (III)	6M HCl, Heating for 25min	C10H8; 535	1.516 x 10 ⁴	Heating on boiling water bath for 25min.	120

Table 4b: Spectrophotometric Determination of PGMs using Hydrazone derivatives as Ligand

Derivative used	Metal Detected	Aqueous conditions; Composition	Solvent; λ _{max} (nm)	Molar Absorptivity (Lmol ⁻¹ cm ⁻¹)	Comments	Reference
	Rh (III)	Buffer, pH 5.5, 20-25 min. digestion time, equilibration time of 1min.; 1:3	CCl ₄ ; 480	8248.23	20-25 min. digestion on boiling water bath, equilibration time of 1min.	121
	Ir (III)	Buffer, pH 6.5, 30 min. heating in boiling water bath, equilibration time of 1min.; 1:3	CH ₃ (CH ₂) ₄ OH; 460	69930	30 min. heating in boiling water bath for digestion; equilibration time of 1min.	122
N-(o-methoxybenzalde hyde) -2-aminophenol (NOMBAP)	Ru (III)	Buffer, pH 5.0, 15-20 min. boiling on water bath digestion time, equilibration time of 1min.; 1:3	CHCl ₅ ; 490	14177.21	Digestion in boiling water bath for 10-15min.; equilibration time of 1min.	123
2-[(5-bromo-2-pyridylazo)]-5-	Pt (II)	Buffer, pH 5.0±0.2, SnCl ₂ .; 2 min. equilibration time; TX-100; 1:1:1:1	CH ₂ Cl ₂ ; 575	6.0 x 10 ⁴	Equilibration time of 2 min. required	124
diethylamino-phenol (5-Br-PADAP)	Pd (II)	Buffer, pH 3.53; 1:2	CH ₂ Cl ₂ ; 585	3.86 x 10 ⁴	Cr(VI), Fe (II) and Th (IV) interfere	125
	Ru (IV)	Buffer; equilibration time 2 min.; 1:3	CH ₂ Cl ₂ ; 560	3.0 x 10 ⁴	Equilibration time of 2 min. required	126

Figure 6: Hydrazones derivatives as chromogenic reagents

2.4 Miscellaneous

2.4.1 4H-1-Benzopyrans

In medicinal chemistry, benzopyrans (Figure 7) are a special kind of bicyclic heterocyclic systems. The prefix "benzo" in the name denoted a ring of benzene fused with a structure of pyran. Any heterocyclic ring containing an oxygen atom and carbons is referred to as a "pyran." Both natural and synthetic products often contain the benzopyran ring system [127]. Natural and synthetic drug building blocks are primarily composed of pyran heterocyclic compounds [128-132]. Heterocyclic compounds containing pyran were categorized according to the hydrogen atom's location and place of origin. The 4*H*-1-benzopyran derivatives find a large scope in the field of research involving material, medicinal, agriculture and biological chemistry [133]. From a survey on literature, 4*H*-1-benzopyrans (Figure 8) are found to possess a good chromogenic and chelating

tendency, hence proving as a versatile class of complexing agents that are used for the spectrophotometric determination of the metal ions including PGMs. The groups responsible for their analytical/chelating behaviour are the oxygen donor carbonyl and the hydroxyl groups present in their structures [127]. A till date study on the said applicability of the 4*H*-1-benzopyrans has been summarized below:

Figure 7: Benzopyrans

A chloroform extractable, coloured complex of 3-hydroxy-2-tolyl-4H-chromen-4-one (HToC) is formed with platinum in its divalent oxidation state in NaHCO₃ medium. The Pt (II)-HToC complex exhibited absorption maximum at 520 nm. Limit of detection and apparent molar absorptivity of the complex are 0.0147μg mL⁻¹ and 6.790 x 10⁴ L mol⁻¹ cm⁻¹, respectively [134]. The method had been successfully used for analysis of platinum in a wide variety of analytical samples. HToC has also been successfully used to analyze iridium in trivalent state [135]. Satisfactory analysis of various synthetic samples and the radical scavenging potential of the newly developed Ir(III)-HToC complex were the practical applications of the studied method.

Complexation of platinum (II) and iridium (III) was carried out with 3hydroxy-2-(4-methoxyphenyl)-4H-chromone (HMPC). Where the Pt (II)-HMPC complex [136] was produced from a 2M NaHCO₃ as a reaction medium and after its extraction into dichloromethane with the maximum absorption noted at 507 nm, the Ir (III)-HMPC complex [137] was formed in presence of H₃PO₄ medium in the water phase only and exhibiting a maximum absorption at 425 nm. The Pd (II)-HMPC complex showed the molar attenuation coefficient of the two complexes, respectively, was 7.1206×10⁴ L mol⁻¹ cm⁻¹ and 6.824 x 10⁴ L mol⁻¹ cm⁻¹. A spectrophotometric method for detecting micro amounts of palladium in its +2 oxidation state had been developed, utilizing a new 1-benzopyran derivative. Using this method, a 1:2 complexation of palladium is made with 3hydroxy-2-(4-methoxyphenyl)-4-oxo-4H-1-benzopyran resulting [Pd(II):HMPB] light yellow extractable complex that shows absorption maximum at 415 nm. The technique satisfied the requirements for accuracy, sensitivity and precision with a molar absorptivity of 3.011 x 10⁴ L mol⁻¹ cm⁻¹ [138].

N. Kaur *et al.* utilized the fluid extraction technique to examine the effect of chromogenic ligand, 3-hydroxy-2-[2'-(5'-methylthienyl)]-4-oxo-4*H*-1-benzopyran (HMTB) on palladium metal in its bivalent oxidation state. With linearity up to 1.5 μ g mL⁻¹, the study demonstrated optimal settings for the complexation of Pd (II)-HMTB in the stoichiometric ratio of 1:2 .This fast and accurate approach works well with a wide range of alloy samples, tailored mixes and industrial goods [139].

A non-extractable light yellow coloured complex of 3-hydroxy-2-(3'-methyl-2'-thienyl)-4-oxo-4*H*-1-benzopran (HMTB) is formed with iridium (III) in phosphoric acid medium at 415 nm. Beers's law obedience of the complex was valid up to 1.5μg mL⁻¹ of Ir (III) and the apparent molar absorptivity as evaluated was 8.45 x 10⁴ L mol⁻¹ cm⁻¹ [140]. Similarly, 6-chloro-3-hydroxy-7-methyl-2-(2'-thienyl)-4-oxo-4*H*-1-benzopyran (CHMTB) formed a yellow complex with iridium (III) in phosphoric acid medium of pH 4.63. Molar extinction coefficient of the complex was 1.18 x 10⁵ L mol⁻¹ cm⁻¹ at 415nm [141]. Palladium in traces had been analyzed in its divalent oxidation state, spectrophotometrically using 6-chloro-3-hydroxy-7-methyl-2-(2'-thienyl)-4-oxo-4*H*-1-benzopyran (CHMTB) as a colouring agent [142]. The metal interacted with CHMTB in a ratio of 1:1 to give a dark yellow complex in NaHCO₃ solution ranging from 0.01-0.025 M strength.

Another benzopyran derivative, 6-chloro-3-hydroxy-7-methyl-2-(2'-furyl)-4*H*-chromen-4-one (CHMFC), helped in determining Palladium in divalent state from a basic medium provided by 0.025-0.040 M sodium bicarbonate solution and producing a 1:1 Pd(II)-CHMFC complex [143]. The method is applied effectively in Pd-charcoal catalyst, alloys, water samples and numerous synthetic mixtures.

Figure 8: Benzopyran derivatives as chromogenic reagents.

2.4.2 Dyes

Colour is a popular attraction for humans and dyes are the coloured compounds that absorb electromagnetic radiations. These compounds are aromatic delocalized electron systems, responsible for absorbing varying wavelengths of radiation. Chromophores, which make energy changes in the delocalized electron cloud of a dye, make it proficient in absorbing radiation. The human eye detects this absorption and responds to colors. However, removal of electrons may cause colour loss or revert to local orbits. This results in an idea of using the dyes as chromophores in the complexation and spectrophotometric detection of the metal ions [144] especially the PGMs. The dyes that had been satisfactorily studied for the metal's analysis are shown in Figure 9.

5-[2-(4-Hydroxyphenyl)hydrazineylidene]-4-iminothiazolidine-2-one, HPIT (Figure 9), in basic medium (2M NaOH) gives coloured species with iridium (IV) which absorbs at 328 nm. The molar absorption coefficient of the complex in the aqueous phase was observed to be 5.57 x 10³ L mol⁻¹ cm⁻¹ [145].

Iridium (IV) gives a coloured complex with Xylene cyanol FF (Figure 9) in the presence of 0.02 M sulphuric acid in the aqueous phase. The complexation required heating at 95°C [146]. A non-extractable species of platinum (IV) is formed with Xylene cyanol FF in sulphuric acid medium. The complex attained maximum colour intensity only after heating at 90°C for 30 minutes. Molar absorptivity of the complex formed was 5.1 X 10⁴ L mol⁻¹ cm⁻¹ at 620 nm, the wavelength of maximum absorption [147].

Investigations were conducted into the Os (IV)-Orange G (Figure 9) complex formation. The reaction's ideal circumstances for the complexation were pH = 5.80~(0.20~M acetate buffer solution); 30 minutes of boiling water bath heating and two-fold reagent excess. At 540 nanometers, the effective molar absorptivity was $1.1~x~10^4~L~mol^{-1}~cm^{-1}$ [148].

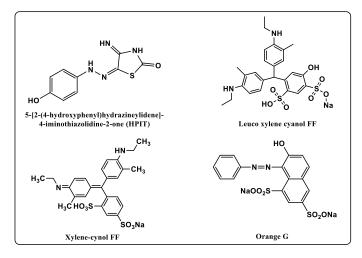


Figure 9: Dyes as chromogenic reagents.

2.4.3 Thiones

Ruthenium (III) forms coloured complexes with BHMBT (Figure 10) in HCl solution with a heating of 7 minutes [149]. The complex formed is extractable in methyl isobutyl ketone (MIBK). The molar absorptivity and λ_{max} have been found to be 5.054×10^3 L mol $^{-1}$ cm $^{-1}$ and 660 nm, respectively. A coloured complex of osmium (VIII) is formed with BHMBT in 1M HCl and 1M HClO4 acid solutions which can be extracted into MIBK with λ_{max} 520 nm and 540 nm, respectively. The molar absorptivity in both the cases has been found to be 4.907×10^4 L mol $^{-1}$ cm $^{-1}$ [150]. At 370 nm, BHMBT is used for the spectrophotometric determination of palladium (II) as its coloured complex in 0.7-3.5M HClO4 solution. The complex is extractable into methyl isobutyl ketone and shows maximum absorption at 370 nm with molar absorptivity 1.543×10^4 L mol $^{-1}$ cm $^{-1}$ [151].

Certain triazoles are also noticed to bind to some PGMs and hence helping in their analysis. One of the triazoles, 4-(4'-fluorobenzylideneamino)-3-methyl-5-mercapto-1,2,4-triazole (FBIMMT; Figure 10) reacts with ruthenium (III) in an acetate buffer medium to give a colored complex between the two. The complex is extractable in n-butanol with maximum absorbance measured at 394 nm. Molar absorptivity of the complex formed is 2.75 x 10³ L mol⁻¹ cm⁻¹ [152]. Similarly, a 1:1 complex of palladium (II) is formed with FBIMMT at room temperature in HCl medium absorbing maximum at 390 nm. The calculated molar absorptivity of the complex is 5.404 x 10³ L mol⁻¹ cm⁻¹ [153].

At pH 5.0, 1, 3-bis (hydroxymethyl) benzimidazole-2-thione (BTAHQ; Figure 10) forms a complex with iridium (III) in aqueous phase with absorbance measured at 563 nm. The apparent molar absorptivity of the complex was evaluated to be 4.81×10^4 and 4.26×10^7 L mol $^{-1}$ cm $^{-1}$ [154].

Figure 10: Thione derivatives as chromogenic reagents

CONCLUSION

The remarkable chemical stability of platinum group metals (PGMs) under diverse environmental conditions and high temperatures makes them indispensable in the fields of chemical engineering and chemistry. Across a wide range of industries, their distinctive qualities make them extremely valuable material. However, as PGM reserves gradually disappear, recycling waste that contains PGMs is imperative in order to guarantee the prudent use of available resources. When it comes to obtaining these metals, recycling catalytic waste, as

opposed to the traditional method of mining from natural ores, offers a more economical and environmentally responsible choice. Recent research indicates that the platinum, palladium, and rhodium group metals (PGMs) that are emitted from car catalytic converters are accumulating beside roadsides. Concerns regarding the toxicity of PGMs to living things and their effects on the environment are being raised by the rising concentrations of PGMs in the environment as a result of this deposition. The growing interest in the medicinal inorganic chemistry, based upon the use of metal complexes especially the PGM complexes as drugs, has led to the detection of metal ions in trace quantities. This review flourishes the arena of spectrophotometric determination of metal ions in their complexes. UV/VIS spectrophotometric technique is having a great demand in wide array of sectors, even though it is regarded as obsolete by the analysts. The fact behind this is the better detection limit and selectivity of the atomic absorption spectrophotometry and the inductively coupled emission spectroscopy along with the mass spectrophotometry. However, still UV-VIS spectrophotometry is well known and used due to its advantages of being inexpensive, easy availability, comparable sensitivity, selectivity and more commonly the accuracy and precision. Advances on the synthesis of different reagents have led the scope for the researchers to work with these techniques for the determination of trace metal ions. The sensitivity and utility of the complexes can be improved using different complexing reagents. Thus, this technique can be regarded as a competitive technique for the detection of metals in atleast trace amounts.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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